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#### REFERENCES

- 1. Anderson, R. M., J. Santanek, and J. C. Harris, JAOCS 37, 119-121 (1960).
- (1960). Ehrenkranz, F., Soap and Chem. Specialties 32, 41-42 (1956). Ginn, M. E., E. L. Brown, and J. C. Harris, JAOCS 38, 361-367 Glutt, M. D., Z. Z. L. J. (1961).
   Ginn, M. E., J. C. Harris, *Ibid. 38*, 605-609 (1961).
   Haahti, E., Scand. J. Clin. Lab. Invest. 18, Suppl. 59 (1961).
   Mankowich, A. M., JACCS 38, 589-594 (1961).

- Oldenroth, O., Fette, Seifen, Anstrichmittel 61, 1220-24 (1959).
   8. Powe, W. C., and W. L. Marple, JAOCS 37, 136-138 (1960).
   9. Rothman, Stephen, ed., "The Human Integument," Publ. 54, Am. Assoc. Advan. Sci., Washington, D.C., p. 150.
   10. Wagg, R. E., and C. J. Britt, J. Textile Inst. Trans. 53, T205-meter. T216.

11. Walter, E., Fette, Seifen, Anstrichmittel 61, 188-193 (1959). 12. Walter, E., WASCHEREI-technik und chemie 13, 484-498

(1960).
13. Warth, A. H., "The Chemistry and Technology of Waxes," 2nd ed., Reinhold Publishing Corp., New York, 1956, p. 138.

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# Three New Oilseeds Rich in cis-11-Eicosenoic Acid<sup>1</sup>

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## Abstract

Marshallia caespitosa Nutt. seed oil (family Compositae) contains 44% cis-11-eicosenoic acid, and is the first oil from the Compositae found to contain a high proportion of  $C_{20}$ -monoenoic acid. Seed oils of Alyssum maritimum (L.) Lam. and of Selenia grandis Martin (family Cruciferae) contain 42 and 58% of the same acid, respectively.

The  $C_{20}$  acids from all three oils were obtained in nearly pure form by fractional distillation of the mixed methyl esters by means of a spinning band column. Permanganate-periodate oxidation of the purified methyl esters yielded predominantly nonanoic and undecanedioic acids.

## Introduction

MARSHALLIA CAESPITOSA is a tufted perennial herb, with a flowering scape about 1 ft high, found from Missouri to Texas (1). A member of the Compositae, it is occasionally used for outdoor decorative plantings. Selenia grandis is a winter annual native to Texas and is not known in cultivation. Alyssum maritimum (sweet alyssum), or according to some authors, Lobularia maritima (L.) Desv., is a common lowspreading perennial of the Cruciferae grown as an annual in many rock gardens (1).

Seed of the first two species were collected from the wild by Department botanists; the Alyssum seed was purchased from a commercial seed company which deals in specialty plant materials not widely available. Seeds were stored at 40F from time of receipt at the laboratory until ground for oil extraction.

The first plant material reported to contain cis-11eicosenoic acid was the liquid wax extracted from Simmondsia chinensis (Link) Schneider (jojoba) seed (5). Since then a number of workers have isolated and characterized this C<sub>20</sub>-monoenoic acid from various Cruciferae seed oils (2,7-9,12). The acid has also been found in some fish oils (9) and in certain seed oils of Sapindaceae (4) and of Ranunculaceae (3); one of the former (Cardiospermum halicacabum L.) contains 42%.

The presence of 43.9% cis-11-eicosenoic acid in the seed oil of a composite, Marshallia caespitosa, is reported here. The acid has not previously been known to occur in seed oils of this family in quantities greater than about 1-2%. In addition, the structure of the C<sub>20</sub>-monoenoic acids, reported earlier in Alyssum (41.8%) and Selenia (58.5%) seed oils (10), also has been proved to be *cis*-11- eicosenoic acid. These three new oils are among the richest sources known for this acid.

### Experimental

Oil Extraction. Oil was obtained from ground seeds of the three species by soxhlet extraction with petroleum ether (bp 30-60C). The solvent was removed in a rotary evaporator. Yield of oil was 22.8% for Marshallia, 31.3% for Alyssum, and 18.3% for Selenia.

Preparation of Methyl Esters. Marshallia seed oil (8.90 g) was saponified by refluxing with 1 N ethanolic potassium hydroxide under nitrogen for  $1\frac{1}{2}$  hr. The saponification mixture was extracted with ethyl ether, acidified, and re-extracted with ethyl ether; a yield of 0.48 g of unsaponifiable material and 7.70 g of fatty acid was obtained. The unsaponifiables were removed from this oil because it was very dark colored. The fatty acids (6.8 g) were esterified by refluxing 2 hr with 1% sulfuric acid in methanol; 6.4 g of methyl esters were obtained.

Methyl esters of Alyssum and Selenia oil were prepared by transesterification with 1% sulfuric acid in methanol. The yield of Alyssum esters was 99.5% and of Selenia esters, 94.6%.

Spinning Band Column Distillation. The mixed methyl esters of each of the three seed oils were fractionally distilled through a Nester and Faust spinning band column. Pure methyl erucate (C22-monoene) was used as a chaser in the pot for the distillation of Marshallia and Alyssum esters because of the small amount of material available. Recovery of seed oil methyl esters in the distillate was quantitative. A chaser was not used in the distillation of Selenia esters; therefore the recovery of esters in the distillate was only 74% of the charge weight.

Distillation conditions and the weights of collected fractions are shown in Table I. The vacuum in the apparatus was released each time a fraction was removed from the collector. The column was operated at total reflux for some time before each collection. The head temperatures given in Table I were obtained at total reflux and at the end of the collection.

Gas liquid chromatographic (GLC) analysis of selected fractions indicated that fractions 9, 10, and 11 of Marshallia; 8, 9, and 10 of Alyssum; and 2, 3, and 4 of Selenia could be combined and used for identification of the C<sub>20</sub>-monoene. The GLC analyses of the original oil (methyl esters) and the combined distillate fractions are shown in Table II. Infrared spectra (liquid film on sodium chloride plates) of the combined fractions indicated no trans C=C (10.34  $\mu$ ) absorption in any of the three samples.

Identification of the C20-Monoene. Marshallia caespitosa. A 0.13-g portion of combined fractions 9, 10, and 11 was saponified by the usual procedure giving 0.12 g of a viscous colorless liquid. Infrared analysis showed no trans C=C absorption. After three recrys-

<sup>&</sup>lt;sup>1</sup> Presented at the AOCS meeting, Toronto, Canada, 1962. <sup>2</sup> A laboratory of the Utiliz. Res. & Dev. Div., ARS, U.S.D.A.

TABLE I Distillation of Mixed Methyl Esters

	Marshallia caespitosa			Alyssum maritimum			Selenia grandis		
Fraction	Head temp, C	Press., mm Hg	Wt, g	Head temp, C	Press., mm Hg	Wt, g	Head temp, C	Press., mm Hg	Wt, g
1	$\begin{array}{r} 131-135\\ 135-137\\ 138-140\\ 140-141\\ 141-142\\ 143-144\\ 144\\ 144-145\\ 145-147\\ 147-148\\ 148-153\\ 145-157\\ \end{array}$	$\begin{array}{c} 0.35\\ 0.35\\ 0.30\\ 0.30\\ 0.30\\ 0.30\\ 0.30\\ 0.30\\ 0.30\\ 0.30\\ 0.30\\ 0.20\\$	$\begin{array}{c} 1.630\\ 0.750\\ 0.180\\ 0.200\\ 0.190\\ 0.140\\ 0.150\\ 0.410\\ 0.370\\ 0.230\\ 0.225\\ 0.440\\ 0.270\end{array}$	$\begin{array}{c} 142\\ 142-145\\ 140-142\\ 141\\ 140-142\\ 142-144\\ 145-149\\ 149-150\\ 150-151\\ 151\\ 151-153\\ 153-155\\ 155-162\\ \end{array}$	$\begin{array}{c} 0.90\\ 0.70\\ 0.50\\ 0.30\\ 0.35\\ 0.25\\$	$\begin{array}{c} 0.300\\ 0.525\\ 0.640\\ 0.570\\ 0.990\\ 0.430\\ 0.530\\ 0.480\\ 0.570\\ 0.520\\ 0.520\\ 0.260\\ 0.260\\ 0.370\\ \end{array}$	$\begin{array}{c} 130 - 136\\ 136 - 141\\ 142\\ 142 - 143\\ 143 - 144\\ 154 - 156\\ 156 - 160\\ 160 - 165\\ \cdots \\ \cdots$	0.30 0.30 0.30 0.50 a 0.60 0.70 0.70 	2.885 0.960 0.510 0.450 0.540 0.900 0.820 0.460 
14	153-157	0.20	0.650	162-163	0.25	0,560	· ·		

<sup>a</sup> Pot and heating mantle changed.

tallizations from redistilled acetone at -18C, a product was obtained melting at 21.0-22.5C (Lit. value for cis-11-eicosenoic acid 21C, 23-24C) (9).

Another 0.13-g portion of the combined distillate fractions of Marshallia was subjected to permanganate-periodate oxidation in 60% tert-butyl alcohol (11) at room temperature for 17 hr. The oxidation mixture was worked up in the usual manner, and 0.16 g of semisolid material was obtained.

Methyl esters were prepared from a portion of the

TABLE II GLC Analyses of Methyl Esters of Original Oil and Combined Distillate Fractions (Area percent of methyl ester in mixed methyl esters)

	Mar caes	rshallia pitosa	Aly mar	Issum itimum	Selenia grandis	
Acid	Origi- nal oil	Com- bined fract. 9, 10, 11	Origi- nal oil	Com- bined fract. 8, 9, 10	Origi- nal oil	Com- bined fract. 2, 3, 4
Tetradecanoic	0.1		Traco			
Hevedecenoic	3 3	0.1	30	Trogo		0.1
Hexadecenoic	0.2	0.1	Traco	ITace	0.2	0.1
Octadecanoic	1.7	Traco	5.8	0.7	1.9	Trace
Octadecenoic	167	10	20.2	9.0	1.0	20
Octadecadienoic	21 0	Trace	67	0.7	4.9	0.4
Octadecatrianoic	01.0	ITACO	10.1	0.1	1.0	0.4
Eicosanoic	0.4	0.3	10.2	3.6	Traco	0.2
Eicosenoic	120	05.0	41.8	01.6	595	02.6
Eicosedienoic	1 5	00.0	11.0	51.0	30.5	52.0
Docosanoic	1.5		L Å Š		Traco	0.6
Docosenoic		1.8*			3.3	2.2

<sup>a</sup> From methyl erucate chaser.

oxidation mixture by reaction with boron trifluoride in methanol. GLC analysis of the esters gave the results shown in Table III.

The remainder of the oxidation mixture was triturated with 30-60C petroleum ether. The portion not extracted was a white solid, mp 87–91C. After three recrystallizations at -5C from petroleum ether containing ca. 2% methanol, a product melting at 109-110.5C was obtained. One literature value for undecanedioic acid is 109-110C (6). No further work was done with the nonanoic acid fraction.

Alyssum maritimum. A 0.15-g portion of combined fractions 8, 9, and 10 was saponified and an acid obtained (0.13 g) that showed no trans C=C absorption by infrared. After two recrystallizations from abso-

TABLE III GLC Analyses of Oxidation Products (Area percent of methyl ester in mixed methyl esters)

Acid	Marshallia caespitosa	Alyssum maritimum	Selenia grandis		
Nonanoic	41.8	29.5	38.4		
Octanedioic		0.1	0.3		
Nonanedioic	1.3	2.8	3.4		
Decanedioic	Trace	0.5	0.5		
Undecanedioic	56.2	62.9	56.0		
Tridecanedioic	Trace		1.4		
Hexadecanoic		0.2			
Octadecanoic		Trace			
Eicosanoic	0.6	41			

lute ethanol (solid discarded) and two recrystallizations from redistilled acetone at -18C, this acid melted at 21.5–22.0C. On admixture with the  $C_{20}$ monoene obtained from Marshallia no depression of the melting point was observed.

A 0.12-g portion of the combined distillate fractions was oxidized and 0.10 g of products was recovered in the same manner as those of Marshallia. A boron trifluoride-catalyzed methyl ester preparation of the oxidation products had the composition (GLC analysis) shown in Table III. The insoluble product after trituration with petroleum ether was a white solid which, after two recrystallizations at -5C from 30 to 60C petroleum ether containing ca. 2% methanol, melted at 110.0-111.0C. The melting point of a mixture of the two dibasic acids (from Marshallia and Alyssum) was 109.5–110.5C.

Selenia grandis. A 0.21-g portion of the combined distillate was saponified in the usual manner yielding 0.20 g of an acid that showed no trans C=C absorption by infrared. This acid melted at 21.6-22.0C after three recrystallizations from redistilled acetone at -18C. The melting point was undepressed when mixed with the  $C_{20}$ -monoene obtained from Alyssum.

Oxidation of a 0.20-g portion of the combined distillate yielded 0.22 g of a white semisolid product. GLC analysis of the boron trifluoride-catalyzed methyl ester preparation gave the results shown in Table III.

The remaining fatty acids were triturated in the same manner as the previous oxidation mixtures. The petroleum ether insoluble portion yielded a white solid melting at 110.9-111.6C after two recrystallizations from ethyl acetate-petroleum ether at -18C. This acid produced no change in the melting points when mixed with the dibasic acids similarly obtained from Marshallia and Alyssum.

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## REFERENCES

1. Bailey, L. H., The Standard Cyclopedia of Horticulture, The Macmillan Co., New York, 1939. 2. Bailga, M. N., and T. P. Hilditch, J. Soc. Chem. Ind. (London) 67, 258-262 (1948). 3. Chisholm, Mary J., and C. Y. Hopkins, Can. J. Chem. 34, 459-464 (1956)

- 64 (1956).
   4. Chisholm, Mary J., and C. Y. Hopkins, *Ibid.* 36, 1537-1540
- (1958).
  5. Green, R. A., and E. O. Foster, Botan. Gaz. 94, 826 (1933).
  6. Gunstone, F. D., M. A. McGee, Chem. & Ind. (London), 1112
- Hatt, H. H., and A. Z. Szumer, J. Sci. Food Agr. 4, 273-278
- (1953).
  8. Hopkins, C. Y., Can. J. Research 24B, 211-220 (1946).
  9. Hopkins, C. Y., and Mary J. Chisholm, Can. J. Chem. 32, 1033-1038 (1954)

Hopkins, C. Y., and Mary J. Chisholm, Can. J. Chem. 32, 1033-1038 (1954)
 Mikolajczak, K. L., T. K. Miwa, F. R. Earle, I. A. Wolff, and Quentin Jones, JAOCS 38, 678-681 (1961).
 von Rudloff, E., Can. J. Chem. 34, 1413-1418 (1956).
 Youngs, C. G., T. M. Mallard, B. M. Craig, and H. R. Sallans, *Ibid.* 29, 871-876 (1951).

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